

# Intermediate Bayesian Data Analysis Using WinBUGS and BRugs

ENAR 2007 Tutorial

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*presented by*

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# Basics of Bayesian Inference

- As usual, we start with a **likelihood** (or **model**)  $f(\mathbf{y}|\boldsymbol{\theta})$  for the observed data  $\mathbf{y} = (y_1, \dots, y_n)$  given the unknown parameters  $\boldsymbol{\theta} = (\theta_1, \dots, \theta_K)$

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- Add a **prior** distribution  $\pi(\boldsymbol{\theta}|\boldsymbol{\lambda})$ , where  $\boldsymbol{\lambda}$  is a vector of **hyperparameters**.
- The **posterior** distribution for  $\boldsymbol{\theta}$  is given by

$$\begin{aligned} p(\boldsymbol{\theta}|\mathbf{y}, \boldsymbol{\lambda}) &= \frac{p(\mathbf{y}, \boldsymbol{\theta}|\boldsymbol{\lambda})}{p(\mathbf{y}|\boldsymbol{\lambda})} = \frac{p(\mathbf{y}, \boldsymbol{\theta}|\boldsymbol{\lambda})}{\int p(\mathbf{y}, \boldsymbol{\theta}|\boldsymbol{\lambda}) d\boldsymbol{\theta}} \\ &= \frac{f(\mathbf{y}|\boldsymbol{\theta})\pi(\boldsymbol{\theta}|\boldsymbol{\lambda})}{\int f(\mathbf{y}|\boldsymbol{\theta})\pi(\boldsymbol{\theta}|\boldsymbol{\lambda}) d\boldsymbol{\theta}} = \frac{f(\mathbf{y}|\boldsymbol{\theta})\pi(\boldsymbol{\theta}|\boldsymbol{\lambda})}{m(\mathbf{y}|\boldsymbol{\lambda})}. \end{aligned}$$

We refer to this formula as *Bayes' Theorem*.

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- Since  $\lambda$  will usually not be known, a second stage (**hyperprior**) distribution  $h(\lambda)$  will be required, so that

$$p(\boldsymbol{\theta}|\mathbf{y}) = \frac{p(\mathbf{y}, \boldsymbol{\theta})}{p(\mathbf{y})} = \frac{\int f(\mathbf{y}|\boldsymbol{\theta})\pi(\boldsymbol{\theta}|\boldsymbol{\lambda})h(\boldsymbol{\lambda}) d\boldsymbol{\lambda}}{\int \int f(\mathbf{y}|\boldsymbol{\theta})\pi(\boldsymbol{\theta}|\boldsymbol{\lambda})h(\boldsymbol{\lambda}) d\boldsymbol{\theta}d\boldsymbol{\lambda}} .$$

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- For prediction of a future value  $y_{n+1}$ , we would use the **predictive** distribution,

$$p(y_{n+1}|\mathbf{y}) = \int p(y_{n+1}|\boldsymbol{\theta})p(\boldsymbol{\theta}|\mathbf{y})d\boldsymbol{\theta} ,$$

which is nothing but the posterior of  $y_{n+1}$ .

# Gibbs sampling

- **Gibbs Sampler:** Suppose the joint distribution of  $\boldsymbol{\theta} = (\theta_1, \dots, \theta_K)$  is uniquely determined by the **full conditional distributions**,  $\{p_i(\theta_i | \theta_{j \neq i}), i = 1, \dots, K\}$ .



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- Given an arbitrary set of starting values  $\{\theta_1^{(0)}, \dots, \theta_K^{(0)}\}$ ,

$$\text{Draw } \theta_1^{(1)} \sim p_1(\theta_1 | \theta_2^{(0)}, \dots, \theta_K^{(0)}),$$

$$\text{Draw } \theta_2^{(1)} \sim p_2(\theta_2 | \theta_1^{(1)}, \theta_3^{(0)}, \dots, \theta_K^{(0)}),$$

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- Under mild conditions,

$$(\theta_1^{(t)}, \dots, \theta_K^{(t)}) \xrightarrow{d} (\theta_1, \dots, \theta_K) \sim p \text{ as } t \rightarrow \infty.$$

# Gibbs sampling (cont'd)

- For  $t$  sufficiently large (say, bigger than  $t_0$ ),  $\{\boldsymbol{\theta}^{(t)}\}_{t=t_0+1}^T$  is a **(correlated)** sample from the true posterior.

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- Most popular software package for this: **WinBUGS**
  - Uses **R**-like syntax to specify models
  - freely available from <http://www.mrc-bsu.cam.ac.uk/bugs/welcome.shtml>

# Gibbs sampling (cont'd)

- In practice, we may actually run  $m$  *parallel* Gibbs sampling chains, instead of only 1, for some modest  $m$  (say,  $m = 5$ ). Discarding the burn-in period, we obtain

$$\hat{E}(\theta_i | \mathbf{y}) = \frac{1}{m(T - t_0)} \sum_{j=1}^m \sum_{t=t_0+1}^T \theta_{i,j}^{(t)},$$

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- If the full conditional  $p(\theta_i | \theta_{j \neq i}, \mathbf{y})$  is not available in closed form, it will typically still be available **up to proportionality constant**. So WinBUGS uses:
  - **adaptive rejection sampling** (log-concave densities)
  - **slice sampling** (bounded domains)
  - **Metropolis sampling** (all other cases)



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$$\int_{-\infty}^{q_L} p(\theta|\mathbf{y})d\theta = \alpha/2 \quad \text{and} \quad \int_{q_U}^{\infty} p(\theta|\mathbf{y})d\theta = \alpha/2 .$$

Then clearly  $P(q_L < \theta < q_U|\mathbf{y}) = 1 - \alpha$ ; our confidence that  $\theta$  lies in  $(q_L, q_U)$  is  $100 \times (1 - \alpha)\%$ . Thus this interval is a  $100 \times (1 - \alpha)\%$  **credible set** (“Bayesian CI”) for  $\theta$ .

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- Though not necessarily narrowest, this **equal tail** interval is easy to compute.
- Unlike frequentist CIs, interpretation of Bayesian CIs is direct: **“The probability that  $\theta$  lies in  $(q_L, q_U)$  is  $(1 - \alpha)$ .”**

# Bayesian hypothesis testing

- Classical approach bases accept/reject decision on

$$\text{p-value} = P\{T(\mathbf{Y}) \text{ more "extreme" than } T(\mathbf{y}_{obs}) | \boldsymbol{\theta}, H_0\} ,$$

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- Bayesian approach: for two models, a commonly used summary historically is the **Bayes factor**,

$$BF = \frac{P(M_1|\mathbf{y})/P(M_2|\mathbf{y})}{P(M_1)/P(M_2)} = \frac{p(\mathbf{y} | M_1)}{p(\mathbf{y} | M_2)},$$

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- **Problem:** If  $\pi_i(\boldsymbol{\theta}_i)$  is **improper**, then  $p(\mathbf{y} | M_i)$  necessarily is as well  $\implies$  **BF is not well-defined!**...

# Bayesian hypothesis testing via DIC

- A generalization of the Akaike Information Criterion (AIC) to the case of hierarchical models based on the posterior distribution of the deviance statistic,

$$D(\boldsymbol{\theta}) = -2 \log f(\mathbf{y}|\boldsymbol{\theta}) + 2 \log h(\mathbf{y}) ,$$

where  $f(\mathbf{y}|\boldsymbol{\theta})$  is the likelihood and  $h(\mathbf{y})$  is any standardizing function of the data alone



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- Summarize the fit of a model by the posterior expectation of the deviance,  $\bar{D} = E_{\theta|y}[D]$
- Summarize the complexity of a model by the effective number of parameters,

$$p_D = E_{\theta|y}[D] - D(E_{\theta|y}[\boldsymbol{\theta}]) = \bar{D} - D(\bar{\boldsymbol{\theta}}) .$$

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- DIC can be sensitive to **parametrization** and **“focus”** (i.e., what is considered to be part of the likelihood)
  - $f(\mathbf{y}|\theta)$ : “focused on  $\theta$ ”
  - $p(\mathbf{y}|\eta) = \int f(\mathbf{y}|\theta)p(\theta|\eta)d\theta$ : “focused on  $\eta$ ”

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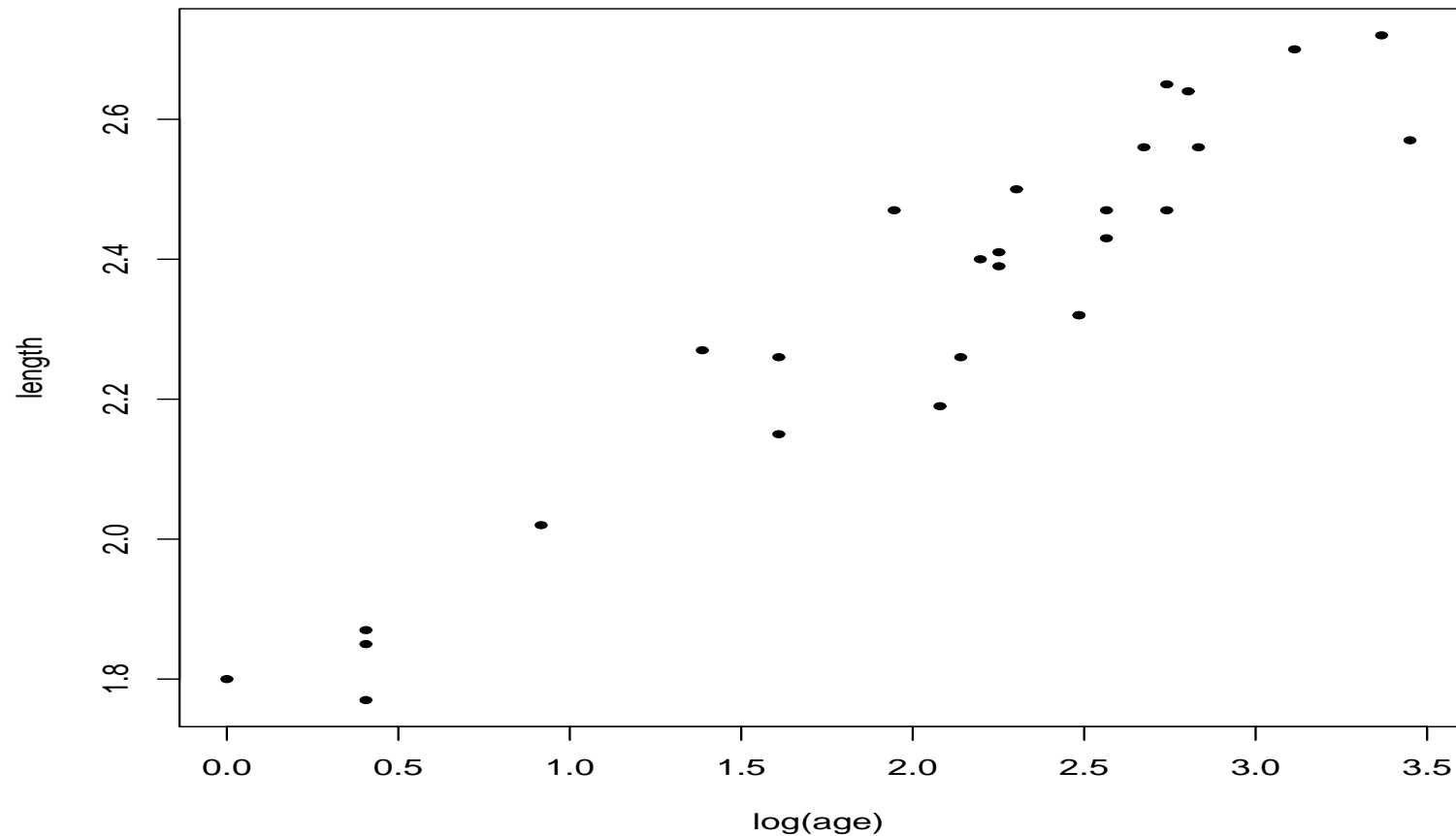
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- Like AIC, DIC tends to select “bigger” models

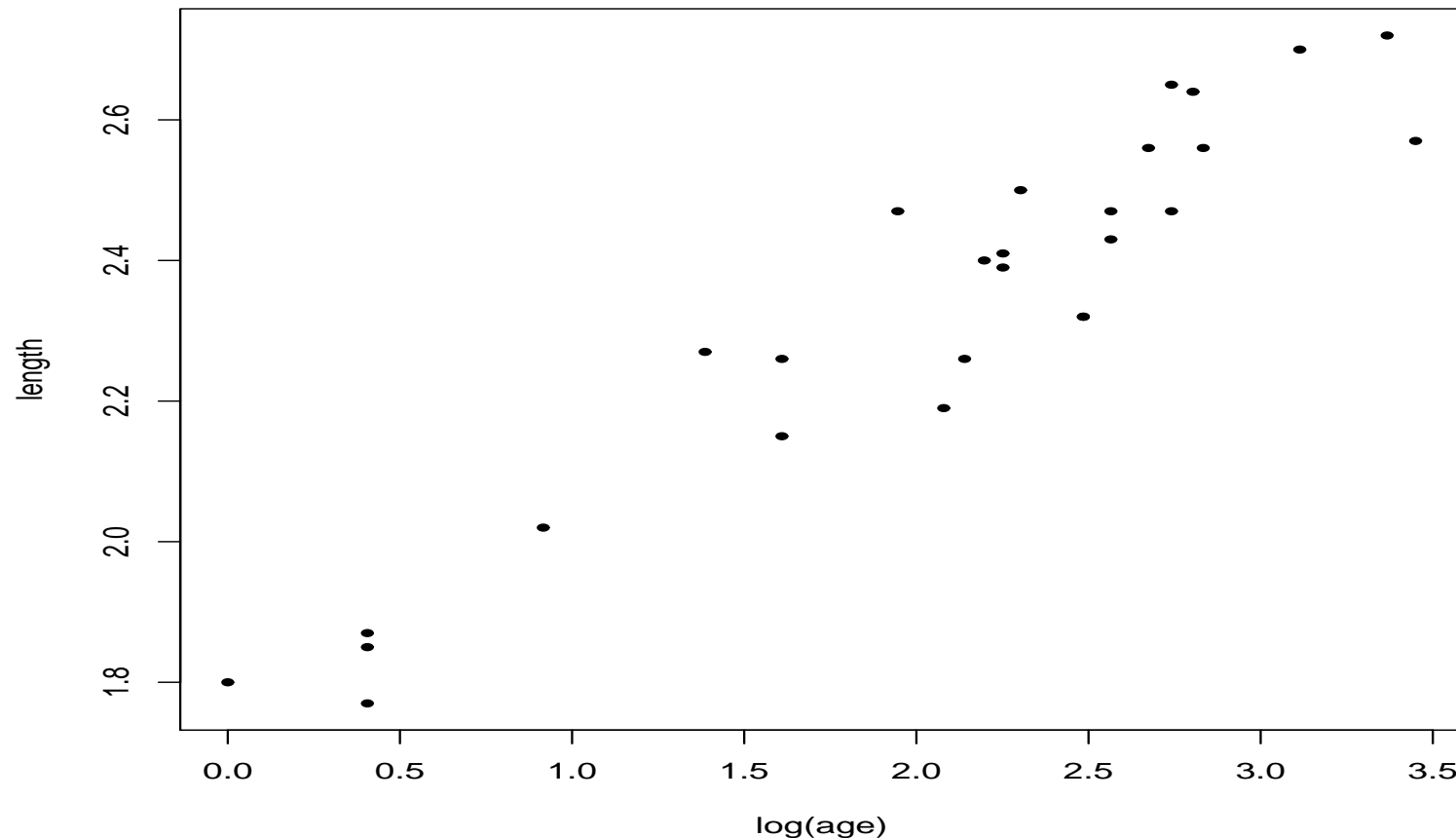
# BUGS Example 1: Linear Regression



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- To avoid a nonlinear model for now, transform  $x_i$  to the log scale; plot of  $Y$  versus  $\log(x)$  looks fairly **linear**!

# Simple linear regression in WinBUGS

$$Y_i = \beta_0 + \beta_1 \log(x_i) + \epsilon_i, \quad i = 1, \dots, n$$

where  $\epsilon_i \stackrel{iid}{\sim} N(0, \tau)$  and  $\tau = 1/\sigma^2$ , the **precision** in the data.

- Prior distributions:

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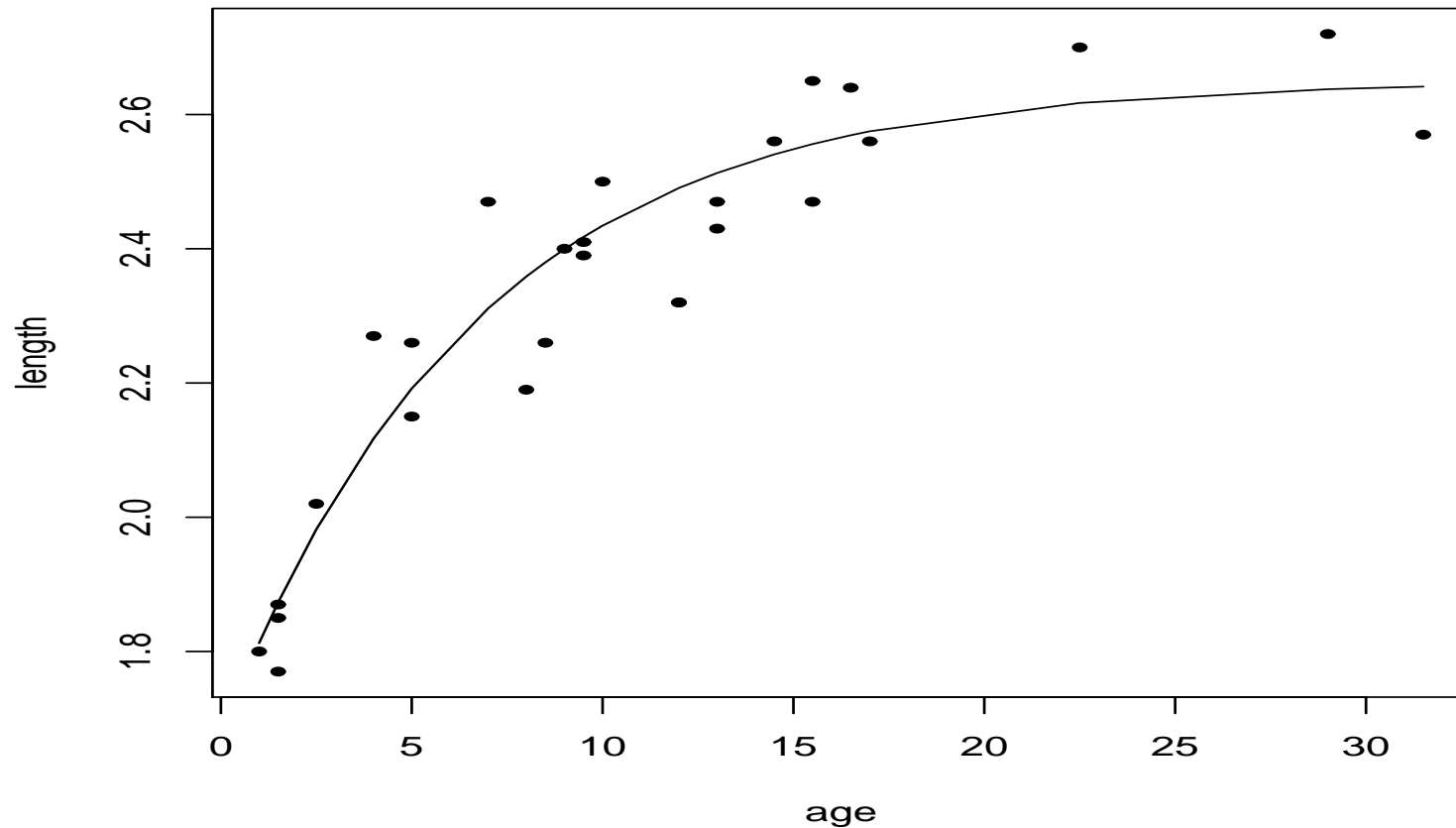
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[www.biostat.umn.edu/~brad/data/dugongs\\_BUGS.txt](http://www.biostat.umn.edu/~brad/data/dugongs_BUGS.txt)

# BUGS Example 2: Nonlinear Regression



- Model the **untransformed** dugong data as

$$Y_i = \alpha - \beta\gamma^{x_i} + \epsilon_i, \quad i = 1, \dots, n,$$

where  $\alpha > 0$ ,  $\beta > 0$ ,  $0 \leq \gamma \leq 1$ , and as usual  $\epsilon_i \stackrel{iid}{\sim} N(0, \tau)$  for  $\tau \equiv 1/\sigma^2 > 0$ .

# Nonlinear regression in WinBUGS

- In this model,
  - $\alpha$  corresponds to the average length of a fully grown dugong ( $x \rightarrow \infty$ )
  - $(\alpha - \beta)$  is the length of a dugong at birth ( $x = 0$ )
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- Obtain posterior density estimates and autocorrelation plots for  $\alpha, \beta, \gamma$ , and  $\sigma$ , and investigate the **bivariate posterior** of  $(\alpha, \gamma)$  using the **Correlation** tool on the **Inference** menu!

# BUGS Example 3: Logistic Regression

- Consider a binary version of the dugong data,

$$Z_i = \begin{cases} 1 & \text{if } Y_i > 2.4 \text{ (i.e., the dugong is "full-grown")} \\ 0 & \text{otherwise} \end{cases}$$

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- A **logistic** model for  $p_i = P(Z_i = 1)$  is then

$$\text{logit}(p_i) = \log[p_i/(1 - p_i)] = \beta_0 + \beta_1 \log(x_i) .$$

# BUGS Example 3: Logistic Regression

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- Two other commonly used link functions are the **probit**,

$$\text{probit}(p_i) = \Phi^{-1}(p_i) = \beta_0 + \beta_1 \log(x_i) ,$$

and the **complementary log-log** (cloglog),

$$\text{cloglog}(p_i) = \log[-\log(1 - p_i)] = \beta_0 + \beta_1 \log(x_i) .$$

# Binary regression in WinBUGS

- Code:  
[www.biostat.umn.edu/~brad/data/dugongsBin\\_BUGS.txt](http://www.biostat.umn.edu/~brad/data/dugongsBin_BUGS.txt)

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model	$\bar{D}$	$p_D$	DIC
logit	19.62	1.85	21.47
probit	19.30	1.87	21.17
cloglog	18.77	1.84	20.61

In fact, these scores can be obtained **from a single run**; see the **“trick version”** at the bottom of the BUGS file!



# Binary regression in WinBUGS

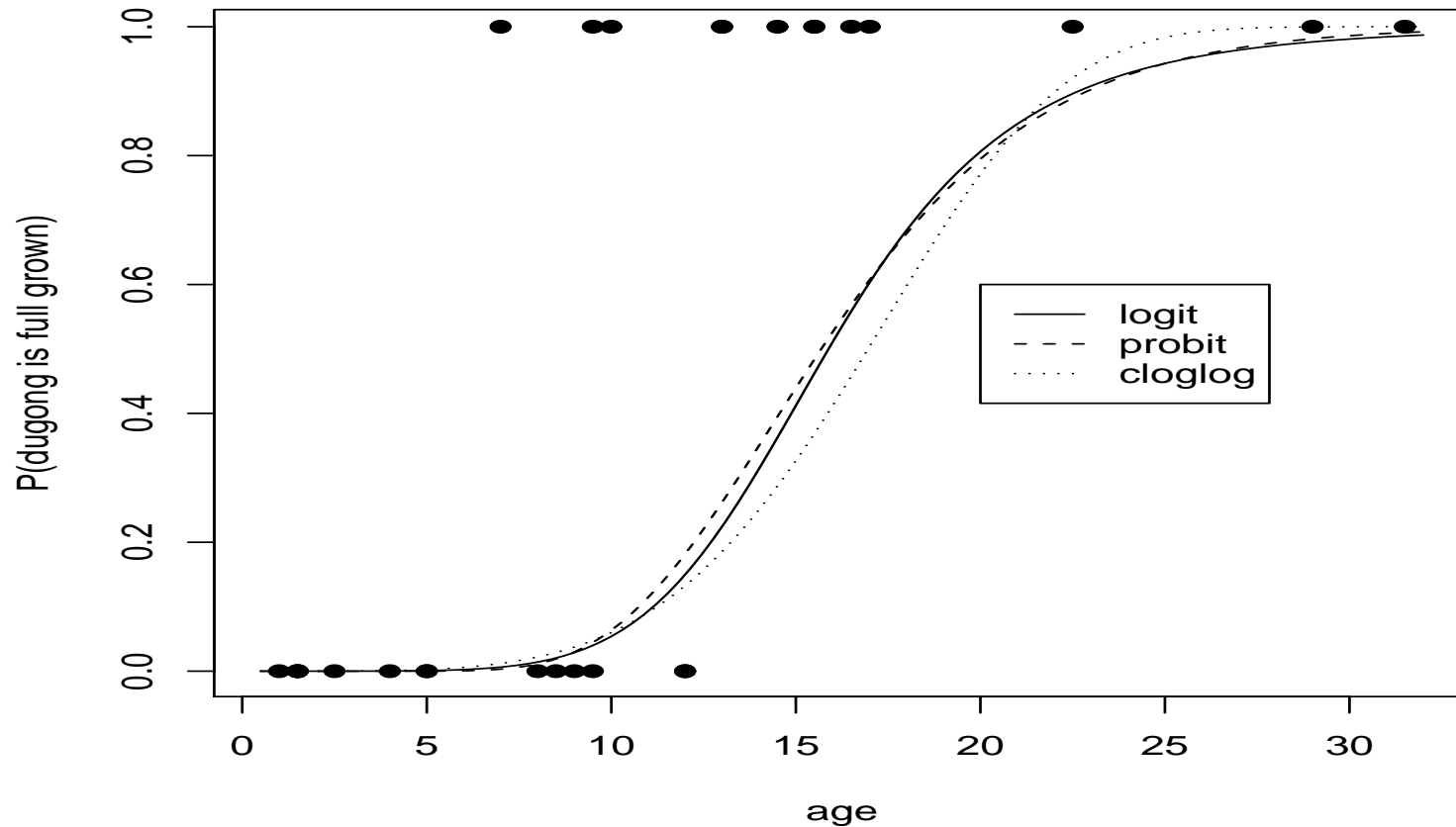
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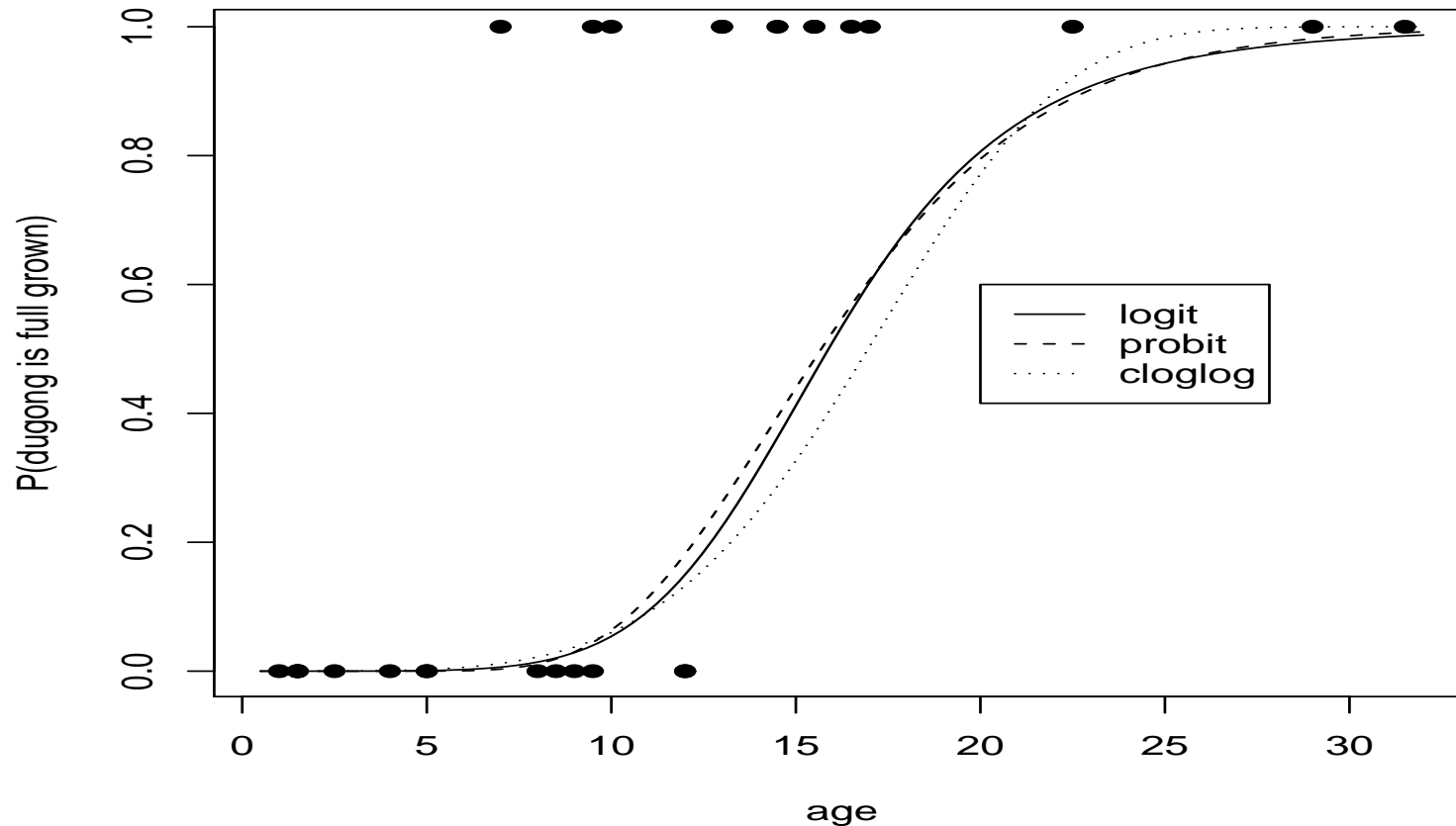
- Use the **Comparison** tool to compare the posteriors of  $\beta_1$  across models, and the **Correlation** tool to check the bivariate posteriors of  $(\beta_0, \beta_1)$  across models.

# Fitted binary regression models



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# Fitted binary regression models



- The logit and probit fits appear very similar, but the cloglog fitted curve is slightly different
- You can also compare  $p_i$  posterior boxplots (induced by the link function and the  $\beta_0$  and  $\beta_1$  posteriors) using the **Comparison** tool.

# BUGS Example 4: Hierarchical Models

- Extend the usual **two-stage** (likelihood plus prior) Bayesian structure to a hierarchy of  $L$  levels, where the joint distribution of the data and the parameters is

$$f(\mathbf{y}|\boldsymbol{\theta}_1)\pi_1(\boldsymbol{\theta}_1|\boldsymbol{\theta}_2)\pi_2(\boldsymbol{\theta}_2|\boldsymbol{\theta}_3)\cdots\pi_L(\boldsymbol{\theta}_L|\boldsymbol{\lambda}).$$

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- $L$  is often determined by the number of **subscripts** on the data. For example, suppose  $Y_{ijk}$  is the test score of child  $k$  in classroom  $j$  in school  $i$  in a certain city. Model:

$$Y_{ijk}|\theta_{ij} \stackrel{ind}{\sim} N(\theta_{ij}, \tau_\theta) \quad (\theta_{ij} \text{ is the } \mathbf{classroom} \text{ effect})$$

$$\theta_{ij}|\eta_i \stackrel{ind}{\sim} N(\eta_i, \tau_\eta) \quad (\eta_i \text{ is the } \mathbf{school} \text{ effect})$$

$$\eta_i|\lambda \stackrel{iid}{\sim} N(\lambda, \tau_\lambda) \quad (\lambda \text{ is the } \mathbf{grand mean})$$

Priors for  $\lambda$  and the  $\tau$ 's now complete the specification!

# Cross-Study (Meta-analysis) Data

- **Data:** estimated log relative hazards  $Y_{ij} = \hat{\beta}_{ij}$  obtained by fitting separate Cox proportional hazards regressions to the data from each of  $J = 18$  clinical units participating in  $I = 6$  different AIDS studies.

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- To these data we wish to fit the **cross-study** model,

$$Y_{ij} = a_i + b_j + s_{ij} + \epsilon_{ij}, \quad i = 1, \dots, I, \quad j = 1, \dots, J,$$

where  $a_i$  = study main effect

$b_j$  = unit main effect

$s_{ij}$  = study-unit interaction term, and

$$\epsilon_{ij} \stackrel{iid}{\sim} N(0, \sigma_{ij}^2)$$

and the estimated standard errors from the Cox regressions are used as (known) values of the  $\sigma_{ij}$ .

# Cross-Study (Meta-analysis) Data

Estimated Unit-Specific Log Relative Hazards						
Unit	Toxo	ddl/ddC	NuCombo ZDV+ddl	NuCombo ZDV+ddC	Fungal	CMV
A	0.814	NA	-0.406	0.298	0.094	NA
B	-0.203	NA	NA	NA	NA	NA
C	-0.133	NA	0.218	-2.206	0.435	0.145
D	NA	NA	NA	NA	NA	NA
E	-0.715	-0.242	-0.544	-0.731	0.600	0.041
F	0.739	0.009	NA	NA	NA	0.222
G	0.118	0.807	-0.047	0.913	-0.091	0.099
H	NA	-0.511	0.233	0.131	NA	0.017
I	NA	1.939	0.218	-0.066	NA	0.355
J	0.271	1.079	-0.277	-0.232	0.752	0.203
K	NA	NA	0.792	1.264	-0.357	0.807
:	:	:	:	:	:	:
R	1.217	0.165	0.385	0.172	-0.022	0.203



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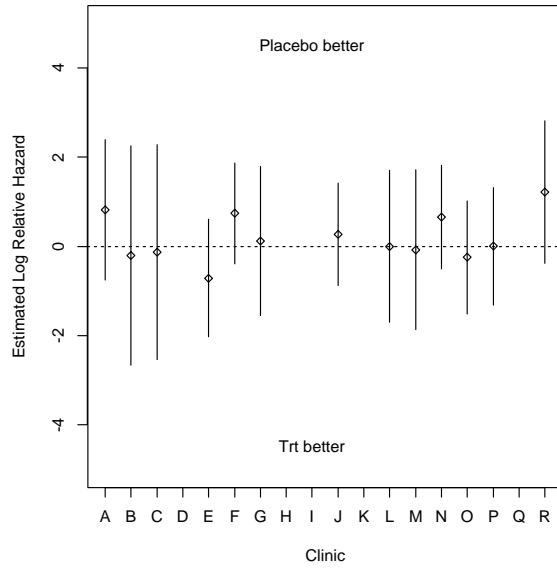
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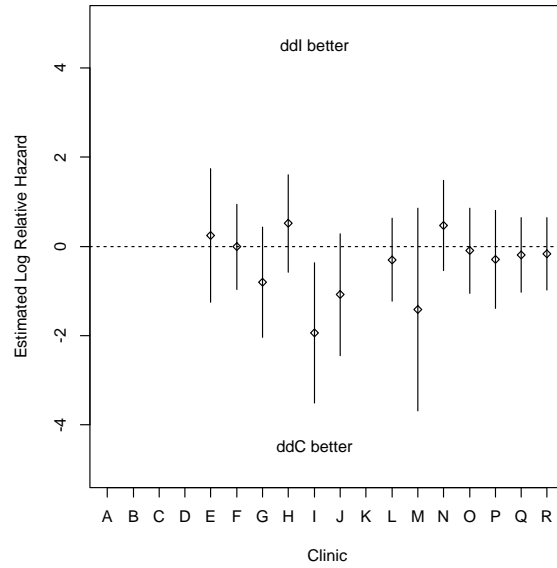
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- Next slide shows a plot of the  $Y_{ij}$  values and associated approximate 95% CIs...

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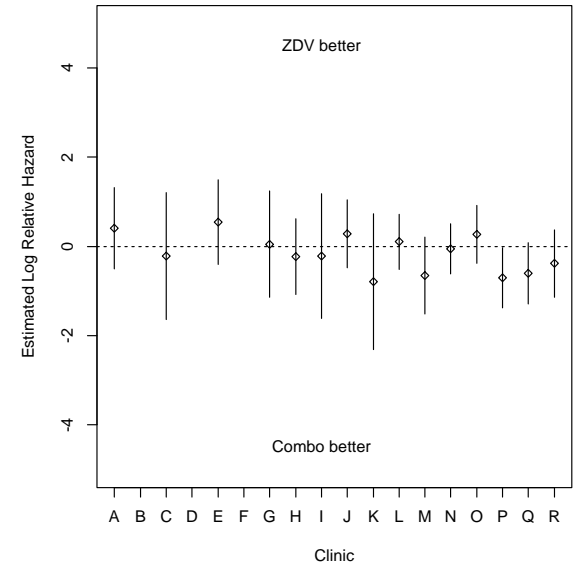
1: Toxo



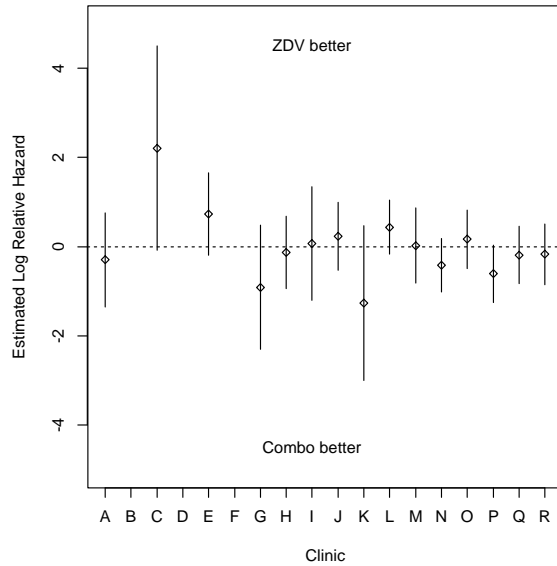
2: ddl/ddC



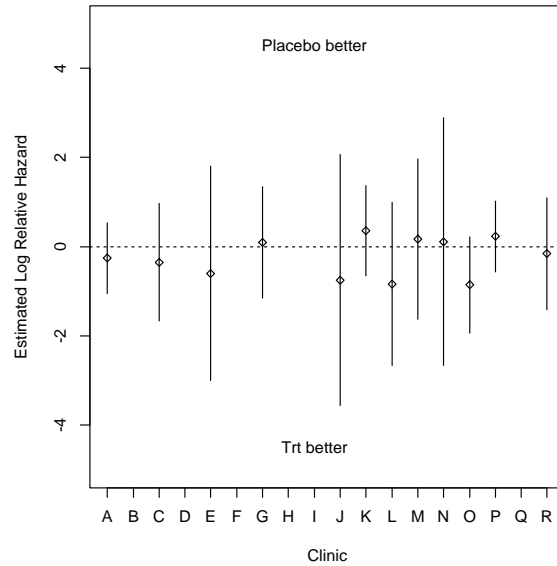
3: NuCombo-ddl



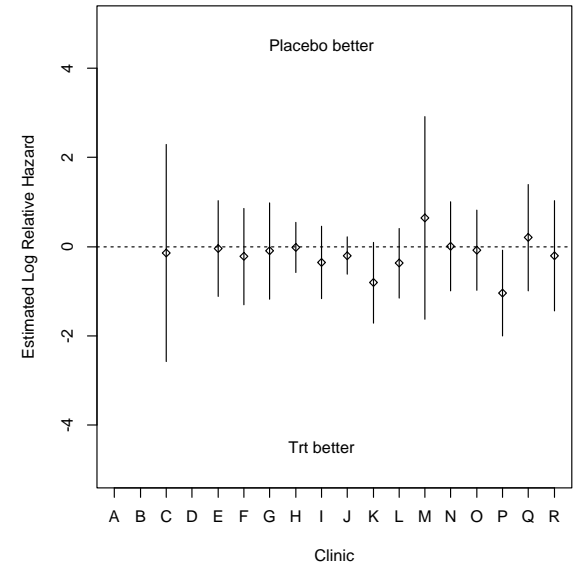
4: NuCombo-ddC



5: Fungal



6: CMV



# Cross-Study (Meta-analysis) Data

- Second stage of our model:

$$a_i \stackrel{iid}{\sim} N(0, 100^2), \quad b_j \stackrel{iid}{\sim} N(0, \sigma_b^2), \quad \text{and} \quad s_{ij} \stackrel{iid}{\sim} N(0, \sigma_s^2)$$

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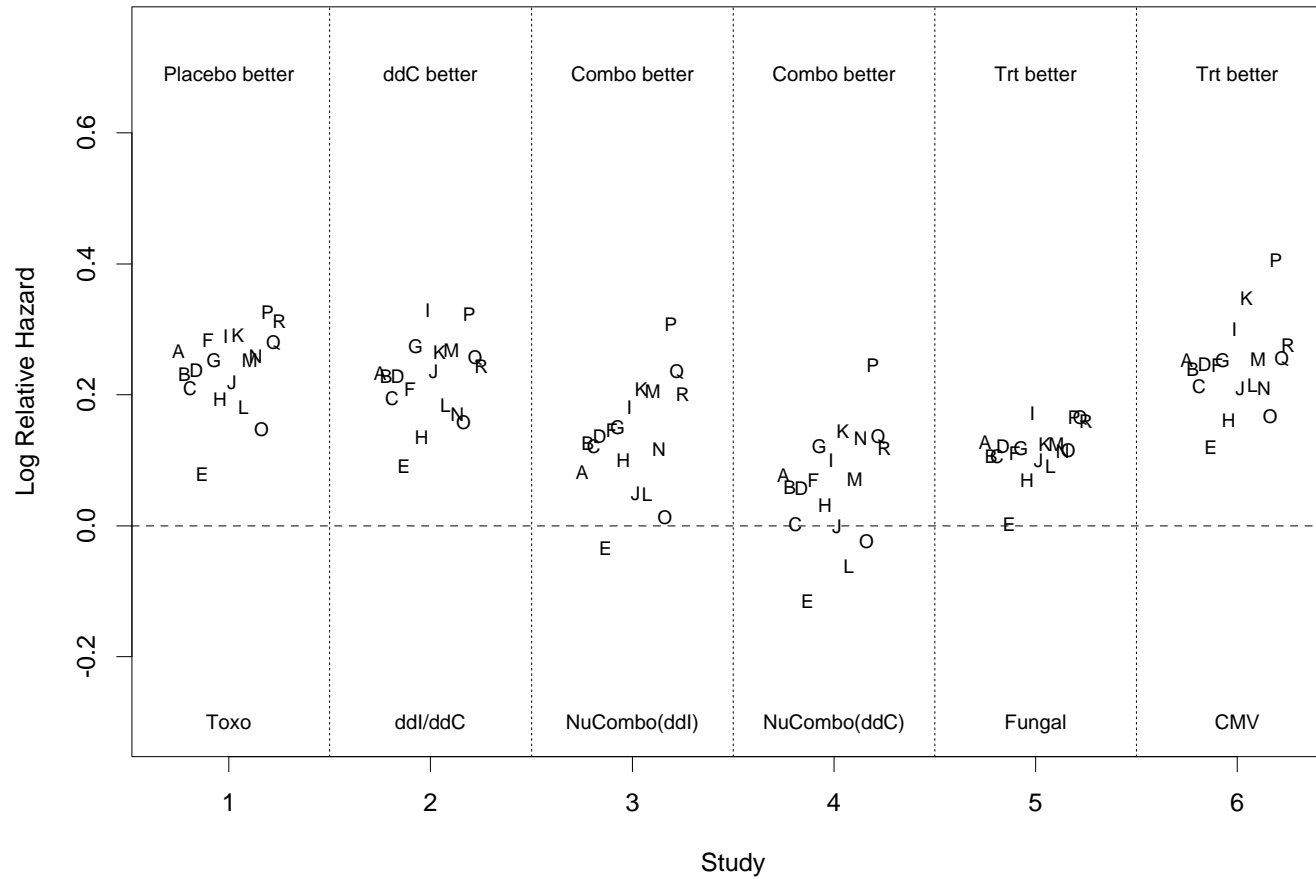
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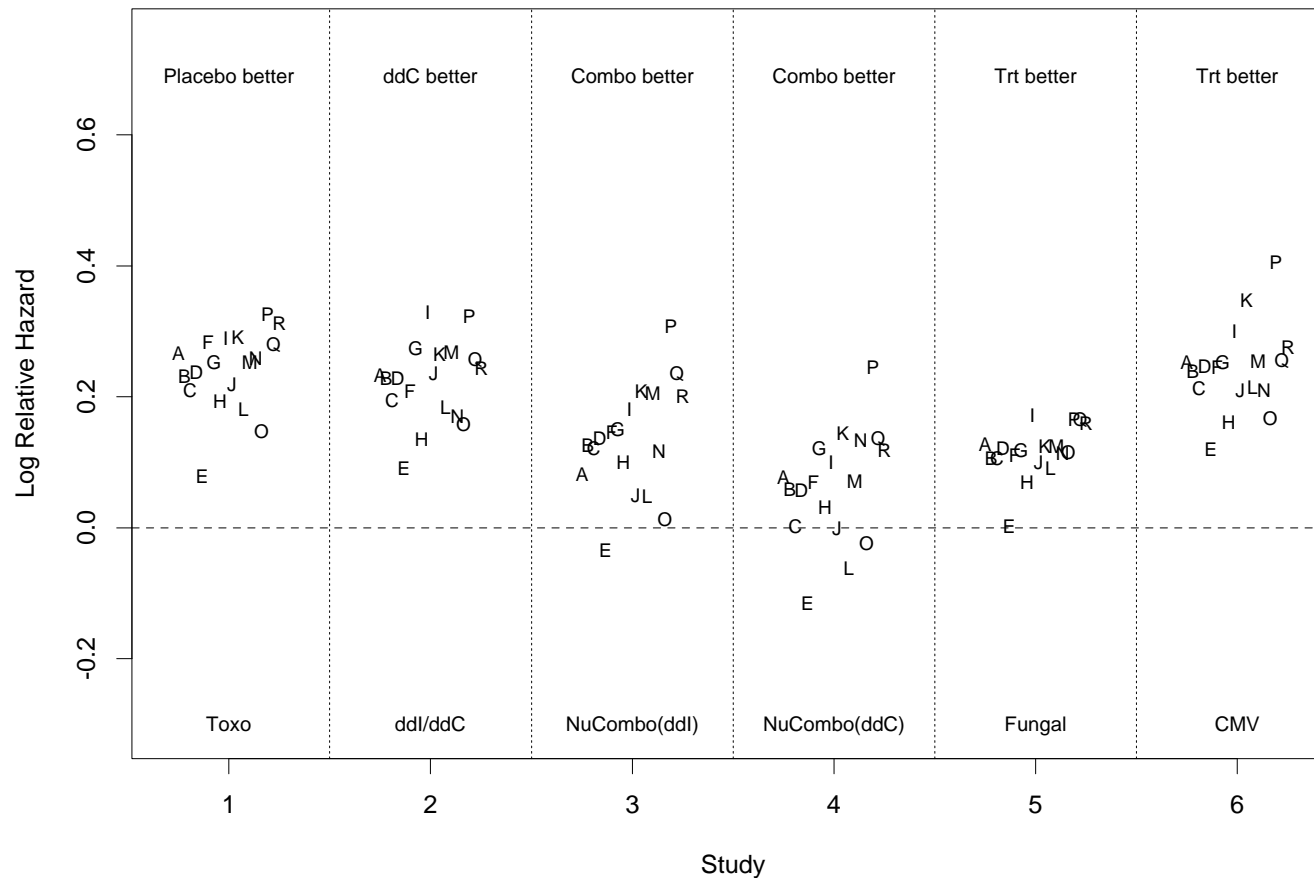
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# Plot of $\theta_{ij}$ posterior means



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# Plot of $\theta_{ij}$ posterior means



- ◇ Unit  $P$  is an opinion leader; Unit  $E$  is a dissenter
- ◇ Substantial shrinkage towards 0 has occurred: mostly positive values; no estimated  $\theta_{ij}$  greater than 0.6

# Model Comparison via DIC

Since we lack replications for each study-unit ( $i$ - $j$ ) combination, the interactions  $s_{ij}$  in this model were only weakly identified, and the model might well be better off without them (or even without the unit effects  $b_j$ ).

As such, compare a variety of reduced models:

```
Y[i,j] ~ dnorm(theta[i,j],P[i,j])
#   theta[i,j] <- a[i]+b[j]+s[i,j]   # full model
#   theta[i,j] <- a[i] + b[j]       # drop interactions
#   theta[i,j] <- a[i] + s[i,j]     # no unit effect
#   theta[i,j] <- b[j] + s[i,j]     # no study effect
#   theta[i,j] <- a[1] + b[j]       # unit + intercept
#   theta[i,j] <- b[j]              # unit effect only
#   theta[i,j] <- a[i]              # study effect only
```

Investigate  $p_D$  values for these models; are they consistent with posterior **boxplots** of the  $b_i$  and  $s_{ij}$ ?

# DIC results for Cross-Study Data:

model	$\bar{D}$	$p_D$	DIC
full model	122.0	12.8	134.8
drop interactions	123.4	9.7	133.1
no unit effect	123.8	10.0	133.8
no study effect	121.4	9.7	131.1
unit + intercept	120.3	4.6	124.9
unit effect only	122.9	6.2	129.1
study effect only	126.0	6.0	132.0

The **DIC-best model** is the one with only an intercept (a role played here by  $a_1$ ) and the unit effects  $b_j$ .

These DIC differences are not much larger than their possible Monte Carlo errors, so almost **any** of these models could be justified here.

# BUGS Example 5: Survival Modeling

- Our data arises from a clinical trial comparing two treatments for *Mycobacterium avium complex (MAC)*, a disease common in late stage HIV-infected persons. Eleven clinical centers (“units”) have enrolled a total of 69 patients in the trial, of which 18 have died.

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- Next page gives survival times (in half-days) from the MAC treatment trial, where “+” indicates a censored observation...



# MAC Survival Data

unit	drug	time	unit	drug	time	unit	drug	time
A	1	74+	E	1	214	H	1	74+
A	2	248	E	2	228+	H	1	88+
A	1	272+	E	2	262	H	1	148+
A	2	344				H	2	162
			F	1	6			
B	2	4+	F	2	16+	I	2	8
B	1	156+	F	1	76	I	2	16+
			F	2	80	I	2	40
C	2	100+	F	2	202	I	1	120+
			F	1	258+	I	1	168+
D	2	20+	F	1	268+	I	2	174+
D	2	64	F	2	368+	I	1	268+
D	2	88	F	1	380+	I	2	276
D	2	148+	F	1	424+	I	1	286+
...	...	...	...	...	...	...	...	...
						K	2	106+

# MAC Survival Data

- With **proportional hazards** and a **Weibull** baseline hazard, stratum  $i$ 's hazard is

$$\begin{aligned}h(t_{ij}; x_{ij}) &= h_0(t_{ij})\omega_i \exp(\beta_0 + \beta_1 x_{ij}) \\ &= \rho_i t_{ij}^{\rho_i - 1} \exp(\beta_0 + \beta_1 x_{ij} + W_i) ,\end{aligned}$$

where  $\rho_i > 0$ ,  $\boldsymbol{\beta} = (\beta_0, \beta_1)' \in \mathbb{R}^2$ , and  $W_i = \log \omega_i$  is a clinic-specific **frailty** term.

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- The  $W_i$  capture overall differences among the clinics, while the  $\rho_i$  allow differing baseline hazards which either increase ( $\rho_i > 1$ ) or decrease ( $\rho_i < 1$ ) over time. We assume i.i.d. specifications for these random effects,

$$W_i \stackrel{iid}{\sim} N(0, 1/\tau) \quad \text{and} \quad \rho_i \stackrel{iid}{\sim} G(\alpha, \alpha) .$$

# MAC Survival Data

- As in the `mice` example (WinBUGS Examples Vol 1),

$$\mu_{ij} = \exp(\beta_0 + \beta_1 x_{ij} + W_i) ,$$

so that

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- Data:** [www.biostat.umn.edu/~brad/data/MAC.dat](http://www.biostat.umn.edu/~brad/data/MAC.dat)  
**Code:** [www.biostat.umn.edu/~brad/data/MACfrailty\\_BUGS.txt](http://www.biostat.umn.edu/~brad/data/MACfrailty_BUGS.txt)

# MAC Survival Results

node (unit)	mean	sd	MC error	2.5%	median	97.5%
$W_1$ (A)	-0.04912	0.835	0.02103	-1.775	-0.04596	1.639
$W_3$ (C)	-0.1829	0.9173	0.01782	-2.2	-0.1358	1.52
$W_5$ (E)	-0.03198	0.8107	0.03193	-1.682	-0.02653	1.572
$W_6$ (F)	0.4173	0.8277	0.04065	-1.066	0.3593	2.227
$W_9$ (I)	0.2546	0.7969	0.03694	-1.241	0.2164	1.968
$W_{11}$ (K)	-0.1945	0.9093	0.02093	-2.139	-0.1638	1.502
$\rho_1$ (A)	1.086	0.1922	0.007168	0.7044	1.083	1.474
$\rho_3$ (C)	0.9008	0.2487	0.006311	0.4663	0.8824	1.431
$\rho_5$ (E)	1.143	0.1887	0.00958	0.7904	1.139	1.521
$\rho_6$ (F)	0.935	0.1597	0.008364	0.6321	0.931	1.265
$\rho_9$ (I)	0.9788	0.1683	0.008735	0.6652	0.9705	1.339
$\rho_{11}$ (K)	0.8807	0.2392	0.01034	0.4558	0.8612	1.394
$\tau$	1.733	1.181	0.03723	0.3042	1.468	4.819
$\beta_0$	-7.111	0.689	0.04474	-8.552	-7.073	-5.874
$\beta_1$	0.596	0.2964	0.01048	0.06099	0.5783	1.245
$RR$	3.98	2.951	0.1122	1.13	3.179	12.05



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- Units C and K have low overall risk ( $W_i < 0$ ) and decreasing hazards ( $\rho < 1$ ): no deaths at all; a few survivors
- Drugs differ significantly: CI for  $\beta_1$  (RR) excludes 0 (1)

# MAC Survival Results

- Units A and E have moderate overall risk ( $W_i \approx 0$ ) but increasing hazards ( $\rho > 1$ ): few deaths, but they occur late
  - Units F and I have high overall risk ( $W_i > 0$ ) but decreasing hazards ( $\rho < 1$ ): several early deaths, many long-term survivors
  - Units C and K have low overall risk ( $W_i < 0$ ) and decreasing hazards ( $\rho < 1$ ): no deaths at all; a few survivors
  - Drugs differ significantly: CI for  $\beta_1$  (RR) excludes 0 (1)
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- **Note:** This has all been for two sets of random effects ( $W_i$  and  $\rho_i$ ), called “Model 2” in the BUGS code. You will also see models having three (adding  $\beta_{1i}$ ), one (deleting  $\rho_i$ ), or zero sets of random effects!

# BRugs Example 1: Model assessment

- Basic tool here is the **cross-validation** residual

$$r_i = y_i - E(y_i | \mathbf{y}_{(i)}) ,$$

where  $\mathbf{y}_{(i)}$  denotes the vector of all the data except the  $i^{th}$  value, i.e.

$$\mathbf{y}_{(i)} = (y_1, \dots, y_{i-1}, y_{i+1}, \dots, y_n)' .$$

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- Also of interest is the **conditional predictive ordinate**,  $p(y_i | \mathbf{y}_{(i)}) = \int p(y_i | \boldsymbol{\theta}, \mathbf{y}_{(i)}) p(\boldsymbol{\theta} | \mathbf{y}_{(i)}) d\boldsymbol{\theta}$ , the height of the conditional density at the observed value of  $y_i$   
 $\implies$  large values indicate good prediction of  $y_i$ .

# Residuals: Approximate method

- Using MC draws  $\boldsymbol{\theta}^{(g)} \sim p(\boldsymbol{\theta}|\mathbf{y})$ , we have

$$\begin{aligned} E(y_i|\mathbf{y}_{(i)}) &= \int \int y_i f(y_i|\boldsymbol{\theta}) p(\boldsymbol{\theta}|\mathbf{y}_{(i)}) dy_i d\boldsymbol{\theta} \\ &= \int E(y_i|\boldsymbol{\theta}) p(\boldsymbol{\theta}|\mathbf{y}_{(i)}) d\boldsymbol{\theta} \\ &\approx \int E(y_i|\boldsymbol{\theta}) p(\boldsymbol{\theta}|\mathbf{y}) d\boldsymbol{\theta} \\ &\approx \frac{1}{G} \sum_{g=1}^G E(y_i|\boldsymbol{\theta}^{(g)}) . \end{aligned}$$



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- Approximation should be adequate unless the dataset is small and  $y_i$  is an extreme outlier
- Same  $\boldsymbol{\theta}^{(g)}$ 's may be used for each  $i = 1, \dots, n$ .

# Approximate methods in WinBUGS

- The ratio to compute the standardized residuals  $d_i$  must be done outside of WinBUGS. Might instead define

$$d_i^* = \frac{y_i - E(y_i|\boldsymbol{\theta})}{\sqrt{Var(y_i|\boldsymbol{\theta})}}.$$

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- For the exact method, we must evaluate  $E(y_i|\mathbf{y}_{(i)})$  and  $Var(y_i|\mathbf{y}_{(i)})$  separately. For the latter, use the facts that  $Var(y_i|\mathbf{y}_{(i)}) = E(y_i^2|\mathbf{y}_{(i)}) - [E(y_i|\mathbf{y}_{(i)})]^2$ , and

$$\begin{aligned} E(y_i^2|\mathbf{y}_{(i)}) &= \int E(y_i^2|\boldsymbol{\theta})p(\boldsymbol{\theta}|\mathbf{y}_{(i)})d\boldsymbol{\theta} \\ &= \int \{Var(y_i|\boldsymbol{\theta}) + [E(y_i|\boldsymbol{\theta})]^2\}p(\boldsymbol{\theta}|\mathbf{y}_{(i)})d\boldsymbol{\theta} . \end{aligned}$$

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- Note that we will now have **both**:
  - an **R** program that organizes the dataset, contains all the `BRugs` commands, and summarizes the output
  - a piece of **BUGS** code that is sent by R to `OpenBUGS`



# Numerical illustration: Stack Loss data

- An oft-analyzed dataset, featuring the stack loss  $Y$  (ammonia escaping), and three covariates  $X_1$  (air flow),  $X_2$  (temperature), and  $X_3$  (acid concentration).

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- WinBUGS code and data for **approximate** method:  
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- See also “**stacks**” in WinBUGS Examples Volume I!

# Approximate vs. Exact Results

obs	sresid		CPO	
	approx	exact	approx	exact
1	0.948	1.098	0.178	0.124
2	-0.566	-0.628	0.224	0.188
3	1.337	1.461	0.122	0.084
4	1.672	1.851	0.078	0.047
5	-0.504	-0.477	0.251	0.244
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- Approximate residuals are too small, especially for the most outlying observations!
- Approximate CPOs also tend to understate lack of fit

# BRugs Example 2: Clinical Trial Design

- Following our MAC survival model, let  $t_i$  be the time until death for subject  $i$ , with corresponding treatment indicator  $x_i$  (= 0 or 1 for control and treatment, respectively). Suppose

$$t_i \sim \text{Weibull}(r, \mu_i), \quad \text{where } \mu_i = e^{-(\beta_0 + \beta_1 x_i)} .$$

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- Then the baseline hazard function is  $\lambda_0(t_i) = r t_i^{r-1}$ , and the median survival time for subject  $i$  is

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- The value of  $\beta_1$  corresponding to a **15% increase in median survival in the treatment group** satisfies

$$e^{\beta_1/r} = 1.15 \iff \beta_1 = r \log(1.15) .$$

# Range of equivalence

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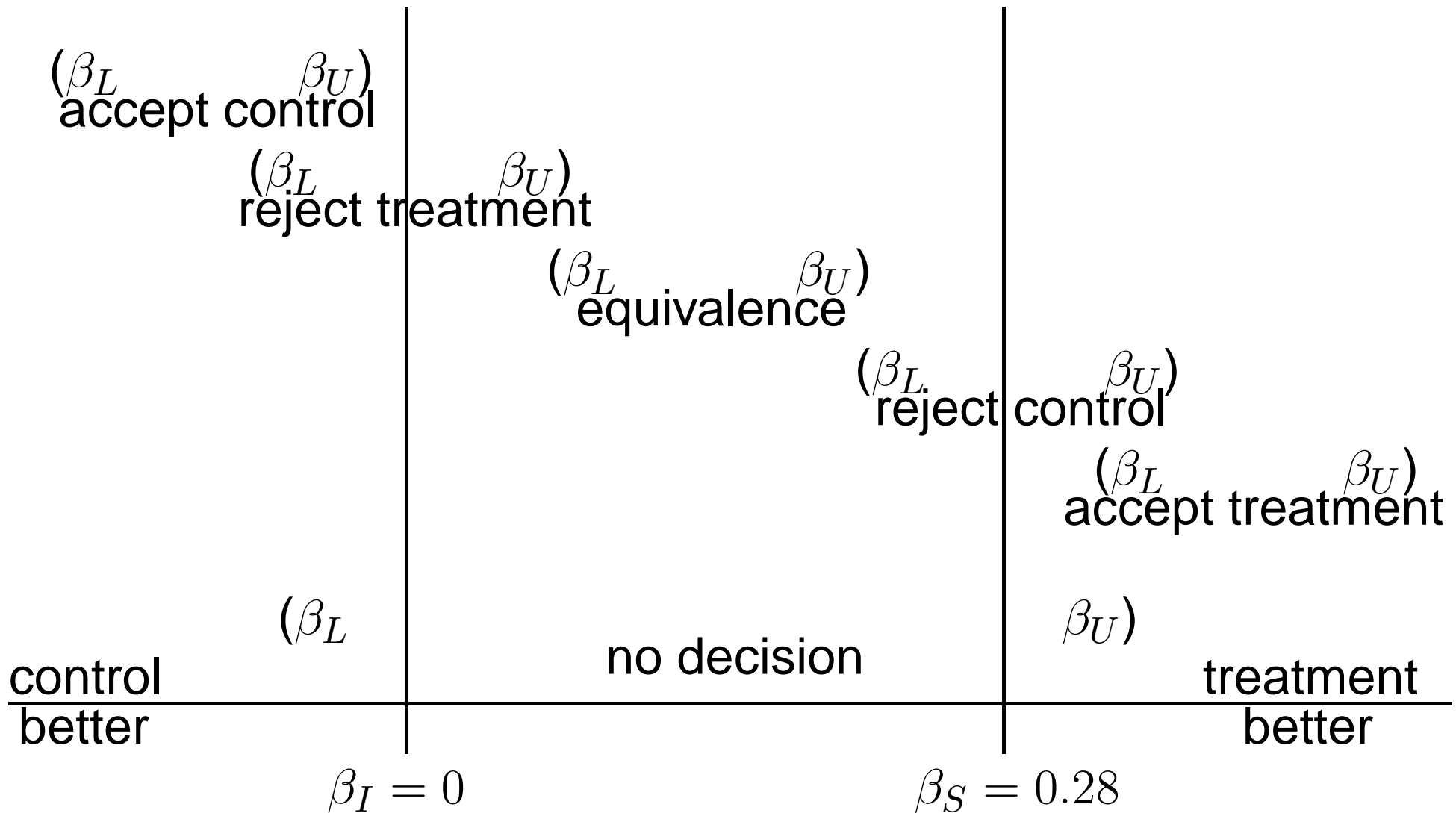
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  - **Example:** If  $r = 2$ , then  $\beta_S = 2 \log(1.15) \approx 0.28$  corresponds to 15% improvement in median survival
- The outcome of the trial can then be based on the location of the 95% posterior confidence interval for  $\beta_1$ , say  $(\beta_L, \beta_U)$ , relative to the indifference zone!....

# The six possible outcomes and decisions



- Note both “acceptance” and “rejection” are possible!

# Community of priors

Spiegelhalter et al. (1994) recommend considering several priors, in order to represent the broadest possible audience:

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- **Reference (or Noninformative)** Prior
  - One that expresses no particular opinion about the treatment's merit
  - Often a **improper uniform** (“flat”) prior is permissible

# MCMC-based Bayesian design

Simulating the power or other operating characteristics (say, Type I error) in this setting works as follows:

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- Repeat this process  $Nrep$  times; report empirical frequencies of the six possible outcomes

# Results from Power.BRugs

- Assuming:
  - Weibull shape  $r = 2$ , and  $N = 50$  in each group
  - median survival of 36 days with 50% improvement in the treatment group
  - a  $N(80, 20)$  censoring distribution
  - the enthusiastic prior as the “truth”

We obtain the following output from  $Nrep = 100$  reps:



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We obtain the following output from  $Nrep = 100$  reps:

- Here are simulated outcome frequencies for  $N = 50$ 
    - accept control: 0
    - reject treatment: 0.07
    - equivalence: 0
    - reject control: 0.87
    - accept treatment: 0.06
    - no decision: 0
- End of BRugs power simulation

# Homework Problems

## ● WinBUGS

- PK hierarchical linear model:  
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● *Thanks for your attention!*